

## Preliminary Investigation for A Potential Ameliorative Role of Methanolic Extract of Leaves of *Polyalthia longifolia* and *Trigonella foenum* and Seeds of *Murraya koenigii* on Lindane Treated Male Rats

Sriram Seshadri<sup>1\*</sup>, Ankit Kumar Jain<sup>1</sup>, Bandish Kapadia<sup>1</sup>

**Abstracts:** Lindane is reported have to detrimental effects on different tissues. In the present study the 90 male Albino Swiss rats were treated lindane at a dosage of 250ppm/kg body weight/day. Following the lindane treatment, shrinkage in the testis was observed but there was no physiological difference in complete body mass. Sperm production was also hampered and the generated sperms were also damaged. Profound histological changes were also observed in the testis and degeneration of the Leydig cells was noted along with reduction in serum testosterone levels. Oxidative stress was also observed to ascent during the lindane treatment course revealing significant generation of ROS. Following the plant extract treatment a significant recovery was observed. The fertility of the rats was restored whereas the functioning of the reproductive organs and accessory sex organs were also apt almost comparable to that of the control group. ROS generation was also observed to decline following the plant extract treatment

**Key Words:** Lindane, *Murraya koenigii*, Oxidative stress, *Polyalthia longifolia*, ROS, Sperm function, Testosterone, *Trigonella foenum*

### INTRODUCTION

Reverse Pharmacology was proposed and initiated by Vaidya. <sup>(1)</sup> Reverse Pharmacology is possible only in those countries with pluralistic healthcare and where robust clinical and laboratory documentation of novel human, pharmacodynamic effects are possible by inter-system collaborative teamwork. <sup>(2)</sup> The scope of Reverse Pharmacology is to understand the mechanisms of action at multiple levels of biological organization and to optimise safety, efficacy and acceptability of the leads in natural products, based on relevant science. <sup>(3)</sup>

Lindane, the  $\gamma$ -isomer of 1,2,3,4,5,6-hexachlorocyclohexane, is a widely used organochlorine pesticide. The application of lindane in soil, foliar, and seed treatment for a large variety of fruit and vegetable crops, its application on livestock, pets, and on agricultural premises, in pharmaceutical preparations, as well as in public health pest control, could lead to the exposure of humans to low concentrations of this compound. Various experiments were carried out in order to assess the toxicity and carcinogenicity of lindane. <sup>(4, 5)</sup> Lindane.  $\gamma$ -HCH intoxication has been associated with male reproductive toxicity in experimental animals and lindane may have the potential to produce adverse effects on fertility in men <sup>(6)</sup> (Kamirin, 1997) as the lindane has the potential of passing through the testis. <sup>(7)</sup> Lindane had also reported to be inducing detrimental effects on various other tissues. <sup>(8, 9)</sup>

Reversal of the reproductive toxicology has already been using vitamin E, D <sup>(10)</sup> and  $\beta$  carotene <sup>(11)</sup> respectively. Treatment with plant extract *Hypericum perforatum* had shown to have the reproductive protective role in metal ions damaged reproductive organs in male mice. <sup>(12)</sup> The antioxidant activities of reproductive parts were higher than those of the vegetative organs, with the pods having highest total phenolic, proanthocyanidin, and flavonoid contents and antioxidant potentials of *Cassia fistula* L. <sup>(13)</sup> However any of the extract had not reported to have proper sperm function and normal function of organs and maturation of the sperm.

The leaves of *Murraya koenigii* are as an herb in Ayurvedic medicine. Their properties include much value as an antidiabetic, antioxidant, antimicrobial, anti-inflammatory, hepatoprotective and anti-hypercholesterolemic. <sup>(14)</sup> *Polyalthia longifolia* had also reported to have cytotoxic property, <sup>(15-18)</sup> Anti-inflammatory properties, <sup>(19)</sup> Antimicrobial properties, <sup>(20)</sup> Hypoglycemic properties. <sup>(21)</sup> *Trigonella foenum-graecum* Fenugreek is a plant in the family Fabaceae has shown many biological prospectuses predominantly includes hypoglycaemic <sup>(22-25)</sup> and cytotoxic properties. <sup>(26-28)</sup> None of the above three plants individually or in combination has been reported to have protective reproductive mechanism. Hence the present study was conducted to understand the toxicological effect of lindane and effect of methanolic extract on the leaves of *Murraya koenigii* and *Polyalthia longifolia* and seeds of *Trigonella foenum-graecum* on the reproductive organs of lindane treated organs.

### MATERIAL AND METHODS

### Experimental Animals

150 healthy adult male rats of Swiss albino strain, weighing between 250g to 300g, were obtained from Dept of Biochemistry, MS University, Vadodara for the experimental purpose. They were housed in an air conditioned animal house at temperature 25 $\pm$ 1 $^{\circ}$ C and exposed to 10 to 12 hours of daylight with free excess to food pellets and water. The Guidance for Care and Use of Animals for Scientific Research [Indian National Science Academy 2000] was strictly followed. The animals were acclimatized for five days prior to the experiments.

### Experiment Schedule

The animals were divided into four groups and caged separately. Group I consisting of 5 animals served as Negative Control Group and was maintained with normal diet and water *ad libitum* through out the study duration. Group II consisted of 10 animals which were given olive oil @ 2 ml/day/rat orally for 90 days study duration. Group III was further divided into 3 sub groups with 45 animals each. These sub groups were fed with lindane dissolved in olive oil at a dosage of 100, 150 and 250 ppm/kg body weight/day respectively for a period of thirty days through a oral gavage attached to a syringe. Group IV comprising of 30 animals from each of Group III sub groups, was fed with 2mL of plant extract per day. Five animals from Group III and IV were sacrificed every ten days in the present investigation, while Group I was sacrificed at the end of 90 day study and Group II was sacrificed at the end of 30 day and 90 days study period.

### Plant Extract

Dried seeds of *Trigonella foenum* were purchased from local dealer. Fresh leaves of *Murraya koenigii* and *Polyalthia longifolia* were collected from Nirma University Botanical garden. The seeds and leaves washed with distilled water. For preparation of extract, air dried powdered seeds of *Trigonella foenum*, leaves of *Murraya koenigii* and *Polyalthia longifolia* was mixed with 60% ethanol in the ratio of 1:10 individually and was soxhlated at 70 $^{\circ}$ C for 72 h. The individual extracts were then mixed in the ratio 2:2:1 to obtain the crude slurry.

### Biochemical Studies

#### Sperm Count and Sperm Viability

The cauda epididymal sperm suspension was prepared in normal saline. The percent motility and count of cauda epididymal spermatozoa of normal and all treated groups of mice were determined by the method of Jeyendra et al (1984) and expressed as percentage motility and millions/ml respectively. The ratio of live: dead

<sup>1</sup>Department of Biochemistry and Biotechnology, Institute of Science, Nirma University, Ahmedabad-382481, Gujarat, India  
Email: sriram.seshadri@nirmauni.ac.in

\*Corresponding author

**Table 1: Experimental Protocol**

Group	Treatment and Dose	Duration (days)	No of animal treated	Day of Autopsy
Group I	Untreated	-----	05	Scarified at the end of 90 days
Group II	Olive oil @ 2 ml/day/rat orally	-----	10	Scarified at the end of 30 days & 90 days
Group III a	Lindane @ 100 ppm/kg body weight/day	30	45	11th, 21st & 31st
Group III b	Lindane @ 150 ppm/kg body weight/day	30	45	11th, 21st & 31st
Group III c	Lindane @ 250ppm/kg body weight/day	30	45	11th, 21st & 31st
Group IV a	Plant extract 2 ml/day/rat orally	60	30	41st, 51st, 61st, 71st, 81st & 91st
Group IV b	Plant extract 2 ml/day/rat orally	60	30	41st, 51st, 61st, 71st, 81st & 91st
Group IV c	Plant extract 2 ml/day/rat orally	60	30	41st, 51st, 61st, 71st, 81st & 91st

**Table 2: Complete Body weight (g) and testis weight (g%/ body weight) of control and treated group animals.**

Group	Complete Body Weight (g)	Testis (g%/ body weight)
Group I	280±4.1	0.46±0.03
Group II	275±3.8	0.45 ±0.02
Group III a	279±0.52	0.43±0.05
Group III b	278±0.39	0.48±0.03
Group III c	277±5.9*	0.37±0.05 <sup>a</sup>
Group IV a	280±0.81	0.44±0.09
Group IV b	279±0.65	0.45±0.04
Group IV c	282±4.9*	0.40±0.07 <sup>a</sup>

Values are mean ± SD. \* no significant <sup>a</sup> P<0.001 Group I = untreated animals, group II= vehicle (olive oil) treated animals; Group III a, b, c = Lindane treated animals for 30 days; group IV a, b, c = Plant extract treated animals for a period of 60 days.

**Table 3: Sperm Analysis and Sperm Functional Analysis of control and treated group animals.**

Parameters	Group I	Group II	Group III a	Group III b	Group III c	Group IV a	Group IV b	Group IV c	
Sperm Analysis	Sperm Count <sup>α</sup>	57.2±4.02	56.4±3.04	55.9±2.76	51.1±4.52	23.32±2.78 <sup>a</sup>	54.8±2.15	56.1±2.05	53.0±2.54 <sup>a</sup>
	Sperm Viability <sup>β</sup>	98.3±2.18	97.9±2.35	94.8±3.87	90.8±2.84	15.4±3.84 <sup>a</sup>	91.2±1.29	90.7±1.41	79.0±1.58 <sup>a</sup>
Sperm Functional Analysis	AIT <sup>γ</sup>	95.4±2.36	97.0±1.58	92.4±5.43	90.7±2.15	18.4±2.07 <sup>a</sup>	91.8±3.16	89.7±2.36	70.6±2.4 <sup>a</sup>
	HOS <sup>γ</sup>	69.2±1.83	68.3±2.12	66.8±3.25	63.0±1.79	22.4±3.50 <sup>a</sup>	70.1±1.95	68.4±1.79	55.0±2.54 <sup>a</sup>
	NCD <sup>γ</sup>	96.3±2.19	96.0±1.87	92.2±3.45	89.4±1.28	28.2±2.38 <sup>a</sup>	91.4±1.48	92.5±3.05	70.8±1.92 <sup>a</sup>

Values are mean ± SD. <sup>a</sup> P<0.001 Group 0 = untreated animals, group I= vehicle (olive oil) treated animals; Group III a, b & c = Lindane treated animals for 30 days; group IV a, b and c = Plant extract treated animals for a period of 60 days. α: million/mL, β: live sperm %, γ: normal %

**Table 4: Seminal Plasma Biochemistry analysis of control and treated animals.**

Parameters	Group I	Group II	Group III a	Group III b	Group III c	Group IV a	Group IV b	Group IV c
Protein <sup>α</sup>	10.23±0.34	11.03±0.28	10.17±0.17	11.2±0.81	7.28±0.56 <sup>a</sup>	9.7±0.41	10.2±0.62	10.34±0.19 <sup>a</sup>
LDH <sup>β</sup>	2.70±0.98	2.77±0.83	2.73±0.20	2.79±0.54	3.57±0.43 <sup>a</sup>	2.75±0.57	2.76±0.31	2.69±0.33 <sup>a</sup>
Acid phosphatase <sup>γ</sup>	20.67±1.68	21.91±1.79	22.01±1.86	21.86±1.46	16.57±1.53 <sup>a</sup>	21.44±1.26	20.97±1.37	18.67±1.87 <sup>a</sup>
Fructose <sup>δ</sup>	22.82±3.16	23.02±3.01	22.17±1.86	22.47±2.15	22.34±3.78 <sup>a</sup>	22.01±1.86	23.04±2.45	21.46±5.90 <sup>a</sup>

Values are mean ± SD. <sup>a</sup> P<0.001, Group I = untreated animals, group II= vehicle (olive oil) treated animals; Group III a, b, c = Lindane treated animals for 30 days; group IV a, b, c = Plant extract treated animals for a period of 60 days. α: mg/100 mg of tissue, β: IU/mg of protein, γ: KA/mg of protein, δ: mM

**Table 5: Tissue biochemical analysis of testis in control and treated animals**

Parameters	Group I	Group II	Group III a	Group III b	Group III c	Group IV a	Group IV b	Group IV c
Total Protein <sup>*</sup>	1.45±0.33	1.40±0.29	1.42±0.19	1.38±0.81	0.82±0.16 <sup>a</sup>	1.32±0.74	1.34±0.66	1.28±0.01 <sup>a</sup>
GSH <sup>α</sup>	224.53±5.08	220.89±4.92	218.7±3.62	215.2±4.76	126.4±6.8 <sup>a</sup>	210.8±4.51	212.5±3.78	210.02±1.05 <sup>a</sup>
MDA <sup>β</sup>	NT	NT	0.03±0.01	0.04±0.02	2.92±0.02 <sup>a</sup>	0.06±0.01	0.05±0.02	0.07±0.02 <sup>a</sup>
Catalase <sup>γ</sup>	35.89±2.05	38.91±3.19	37.3±2.08	36.4±3.54	9.25±0.83 <sup>a</sup>	36.7±3.14	38.2±3.16	37.7±1.37 <sup>a</sup>
SOD <sup>δ</sup>	33.85±1.21	32.73±1.93	32.18±2.05	33.05±0.91	8.73±0.6 <sup>a</sup>	31.7±3.15	32.8±2.57	33.92±0.41 <sup>a</sup>

Values are mean ± SD. <sup>a</sup> P<0.001, Group I = untreated animals, Group II= vehicle (olive oil) treated animals; Group III a, b, c = Lindane treated animals for 30 days; Group IV a, b, c = Plant extract treated animals for a period of 60 days. \*: g%, α: mg/L, β: [10<sup>-5</sup>] (mg/L), γ: H<sub>2</sub>O<sub>2</sub> degraded/min/mg of protein, δ: units/mg of protein

spermatozoa of control and all treated groups of mice were determined using eosin nigrosine staining given by WHO manual. (29) The number of stained and unstained spermatozoa was scored in 10-20 separate fields and percentages (%) were calculated.

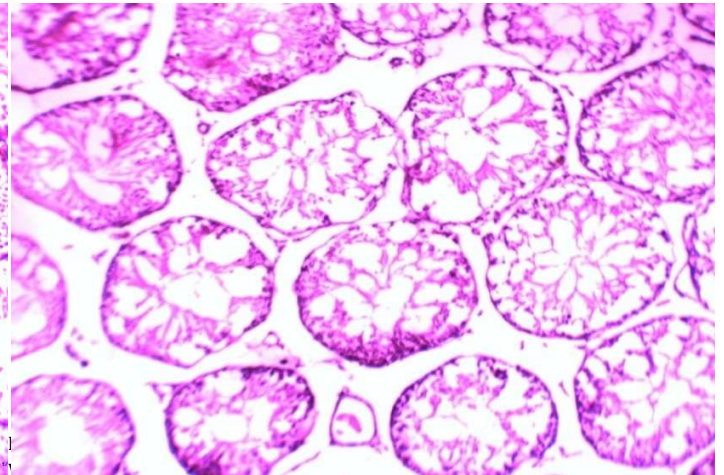
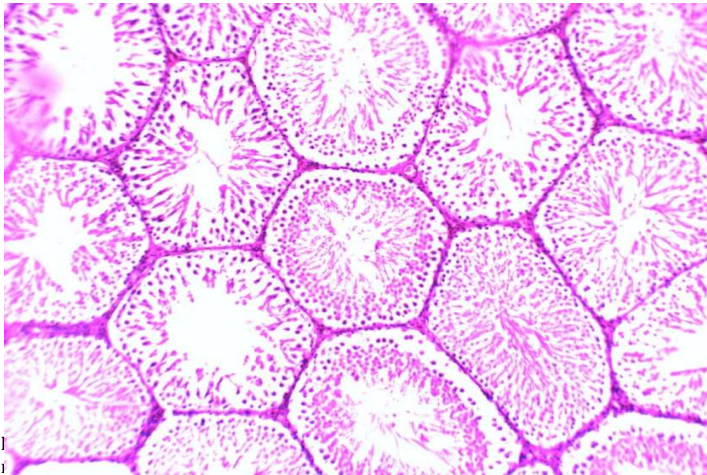
#### Sperm Functional Analysis

Spermatic fluid collected after the tearing of cauda epididymis was used for the assessment of acrosome intactness test (AIT), (30) nuclear chromatin decondensation test (NCD) (31) and Hypo osmotic swelling test (HOS). (32). The score above 50% in all functional test were

considered normal and below the level were considered sub-fertile or infertile.

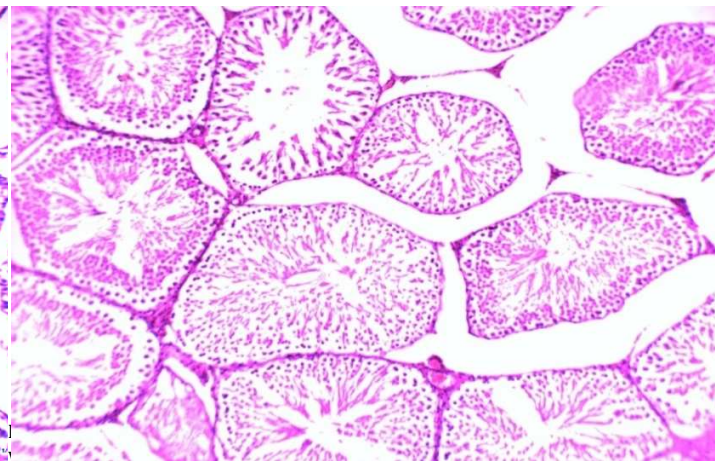
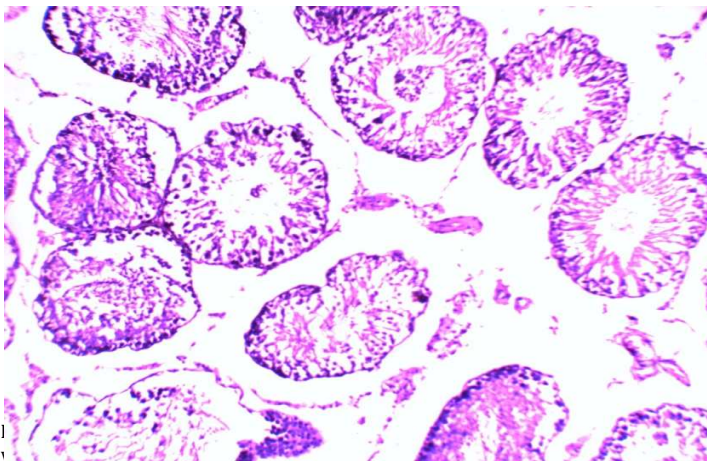
#### Seminal Plasma Biochemistry

Sperm free seminal plasma was used for the quantitative determination of Fructose, (32) Protein, Acid Phosphatase, Lactate



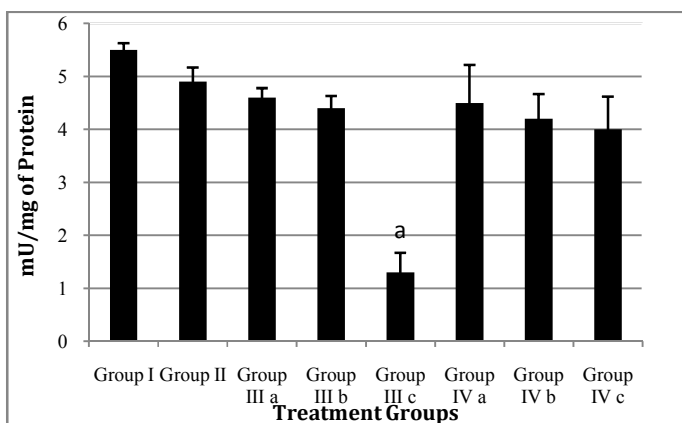
Internal arrangement was observed to be regular, while sperm accumulation (SA) was observed. ST was observed to be connected with interstitial space (IS) where endocrine cells called Leydig cells are located.

Treated Control.



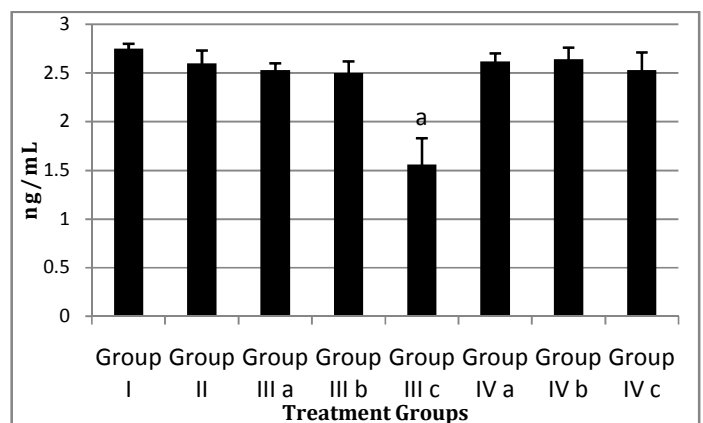
internal arrangement was also completely distorted. Leydig cells were observed to degenerate.

observed to be complete restored while the sperm accumulation was observed in the seminiferous tubules, almost comparable to that of the vehicle treated control animals.



Values are mean ± SD. a: P<0.001  
Group I = untreated animals, Group II= vehicle (olive oil) treated animals; Group III a, b, c = Lindane treated animals for 30 days; Group IV a, b, c = Plant extract treated animals for a period of 60 days.

Figure5: GraphI:α- Glucosidase activity in control and treated animals



Values are mean ± SD. a: P<0.001  
Group I = untreated animals, Group II= vehicle (olive oil) treated animals; Group III a, b, c = Lindane treated animals for 30 days; Group IV a, b, c = Plant extract treated animals for a period of 60 days

Figure 6: GraphII:Testosterone Levels in control and treated animals

Dehydrogenase using (*Qualigen diagnostic kit, Mumbai, India*), and  $\alpha$ -glucosidase (*Roche Applied Science, Germany*).

### Histopathological Studies

Complete testis was removed, cleared of visible fat. The organ weight was taken and a small portion of testis was used for histological examination of control, following the treatment with Lindane and Plant extract. The tissues were cleared of the visible fats and were fixed immediately in Bouin's fluid for 24 hours, dehydrated in ethanol, cleared in benzene, infiltrated and embedded in paraffin wax. Five  $\mu\text{m}$  thick sections were cut and stained with Harris Hematoxylin and eosin.

### Tissue Biochemistry

The remaining tissue was homogenized in phosphate buffer saline, centrifuged at 3500 rpm for the 15 min, the supernatant obtained was used for assaying tissue protein Qualigens Ltd. Mumbai, India, Malondialdehyde (MDA),<sup>(33)</sup> Reduced Glutathione (GSH),<sup>(34)</sup> Super oxide Dismutase (SOD)<sup>(35)</sup> and Catalase<sup>(36)</sup> activities of the vehicle treated control animals, lindane treated and Plant extract treated animals were estimated.

### Testosterone Estimation

Testosterone was estimated using Chemiluminescent Immunoassay Abbott Architect, USA.

### Statistics

For all biochemical estimations, a minimum of 8 to 10 replicates were done for each parameter. The data were statistically analysed using student's t test and Analysis of Variance ANOVA followed by Scheffe's test for least significance. The P value determined was in comparison to control group II.

## RESULT

### Body Weight and Organ Weight

No physiological change in the complete body weight of any of the treated group was observed. Treatment with the lindane, a decline in the testis weight was observed in group III c ( $p < 0.001$ ) after the treatment 30 days in comparison to that of the group I, II, III a and III b. Following treatment with plant extract a gradual recovery was observed in group IV c ( $p < 0.001$ ) almost comparable to that of the control group II. Surprisingly, Group III a and b did not show any significant changes in their body and organ weight (Table 2).

### Sperm Count and Viability

The sperm count in the cauda epididymis of group III c declined significantly ( $P < 0.001$ ) compared other groups. Treated group III a and b showed marginal drop in the sperm count. All the altered values recovered in

group IV animals. The sperm viability data also showed similar trend (Table 3).

### Sperm Functional Analysis

There were no changes observed in any of the sperm functional analysis in any of the study groups expect the Group III c. Group III c showed significant decline in the acrosome intactness test, hypo osmotic swelling test and nuclear chromatin decondensation test suggesting severe functional damage to the sperm. Significant recovery was observed in both HOS ( $P < 0.001$ ) and NCD ( $P < 0.001$ ) within a period of 60 days while AIT also showed gradual recovery comparable to that of the Group II (Table 3).

### Semen Biochemistry

Lindane treatment @ 250 ppm/kg body weight resulted in very significant decrease in seminal plasma biochemical parameters viz., protein, lactate dehydrogenase C<sub>4</sub>, Prostate specific Acid phosphatases activity, Fructose and seminal vesicle specific  $\alpha$ -Glucosidase ( $P < 0.001$ ). while other treated groups ie., Group II, III a and b did not elicit any changes whatsoever. The altered parameters was again restored to almost comparable to that of the control of group I following 60 days of plant extract treatment (Table 4).

### Histopathological Studies

The histology of testis was performed for Group II, Group III b, c and Group IV. The micrograph of testis in control group II demarcates the normal arrangement of the internal structure of the Sertoli cells in the seminiferous tubules. The tubules were observed to be connected through interstitial space containing endocrine cells, Leydig cells. Sperm accumulation was observed in the seminiferous tubules (Figure 1).

Following the lindane treatment @ 150 ppm/kg body weight/day (group III b) for a period of 30 days, did not result in complete degeneration of the interstitial space but there was evidence of initiation of degeneration of Leydig cell. The arrangement of Sertoli cells was also slightly distorted and there was focal degeneration observed. As compared to that Group III c (250 ppm/kg body weight/day) showed significantly distorted seminiferous tubules and in addition, the Leydig cells were completely absent and there was no visible sperm accumulation within the seminiferous tubules. (Figure 2 & 3).

A gradual restoration of the testicular arrangement was observed in animals following a plant extract treatment. During the 30 days of the plant extract treatment (group IV), re-appearance of the connecting tissue was observed in interstitial space indicating regeneration of the Leydig cells. The arrangement Sertoli cells in the seminiferous

tubules were also observed to partially re-arranged compared to that of group II. Sperm accumulation was observed to initiate after 30 days of PE treatment indicating a speedy recovery (Figure not shown).

After the completion of the 60 days of the plant extract treatment the micrograph was almost comparable to that of the vehicle treated control group. The micrograph depicts almost normal arrangement of the interstitial space consisting of connective tissue, Sertoli and Leydig cells were observed to be almost comparable to that of the control group (Figure 4).

### Tissue Biochemistry

The histological observations were confirmed by the tissue biochemical parameters. Following lindane treatment (group III c), a significant decline in the protein content of the tissue was noted. The activity of the oxygen free radical scavenging enzymes (OFR) viz. Super oxide dismutase and catalase were also observed to decline. The concentration of the reduced glutathione was also noted to vary significantly in group III c. In contrast, the levels of the malondialdehyde was noted to escalated while the concentration of malondialdehyde was below the detection limit in Group I, II, III a and b.

Following the plant extract treatment (group IV), gradual recovery was observed in all the altered parameters (Table 5).

### Testosterone

There was a slight non significant drop in the testosterone levels in Group III a and b but Group III c showed a significant decline as compared to the Vehicle treated group. The levels recovered in the plant extract treated animals, almost comparable to that of the control groups (Graph II).

## DISCUSSION

The present study was performed to explore the effects of the lindane and possible ameliorative role of methanolic extract of leaves of *Polyalthia longifolia* and *Trigonella foenum* and seeds of *Murraya koenigii* on the reproductive organs and functions.

Lindane is one of the environmental chemicals known to have an endocrine effect and has been associated with male reproductive alterations.<sup>(37-42)</sup> The mechanism of action of lindane on male reproductive system remains unclear.<sup>(43)</sup> During the lindane treatment, decline in the testicular mass was observed. Similar result were also reported by Beard and Rawling.<sup>(44)</sup> No notable difference in the body mass of the rat was observed during the lindane exposure also reported by Saradha and Mathur.<sup>(43)</sup> However other articles had reported a decline in the body mass following the lindane treatment.<sup>(9)</sup> Thus in the present experiment, shrinkage of

testis was noted but no physiological difference was observed in the complete body mass.

A dropping off in the epididymal sperm count was pragmatic following the lindane treatment which too was also reported by Prasad et al. (39) Sperm viability was also observed to diminish along with damaged sperm. This reveals induction of infertility in male rats due to lindane treatment. An analogous result was also observed by Fausto et al in male rabbits. (45)

In acrosome intactness test (AIT), there was considerable decapitation of the sperm head. Nuclear decondensation (NCD) assay divulged a significant decline in the stability of the chromatin material in the sperm during lindane treatment. Hence the sperm observed to be normal in AIT, but has nuclear instability indicating disturbance in the sperm maturation process. Hypo osmotic (HOS) analysis in lindane treated animal noted to have a significant decline in motility of the sperm. Thus semen analysis and sperm functional analysis reveals that lindane treatment induces infertility and sperm produced are not properly matured viz. sperm produced are damaged, which goes in agreement with Samanta and Chainy. (46)

In the present study, the levels of protein in cauda epididymides showed a significant decrease after 30 days of lindane treatment. This decrease might be due to mutilation of protein metabolism/synthesis.

LDH-C<sub>4</sub>, an isoenzyme associated with testis, routinely used as marker for determining proper functioning of the reproductive organs. (47) An increase in the LDH-C<sub>4</sub> value indicates that the sperm mitochondrial activity has declined; indicating improper functioning of the sperm and maturation. Semen biochemistry analysis showed an increase in activity of LDH-C<sub>4</sub> during lindane treatment, revealing obstruction in the functioning of mitochondria. This may be due to improper maturation or high oxidative stress in the cauda epididymides as reported by Saradha and Mathur. (43)

Acid phosphatase (ACP) and fructose both are associated sperm motility and are secreted by the accessory sex glands. (48). Activity analysis of ACP revealed a decline during lindane treatment, revealing inappropriate environment for motility of the sperm which was also observed during HOS analysis.

The estimation of  $\alpha$ -glucosidase activity in semen is widely studied as marker for epididymal function. (49). Epididymal  $\alpha$ -glucosidase does not play a crucial role in the development of sperm fertilizing capacity, but is involved in the providing optimum condition for sperm storage. (50) In the present study, a dwindle in the activity was noted in

the lindane treated animals, indicating less number of sperm produced as mentioned earlier. The decrease in the enzyme activity has been in observed non-azoospermic men. (51)

Thus treatment of lindane in rats, the functioning of male reproductive organ is complete hampered and the male rats become oligo-necro-zoospermic. Treatment with lindane not only affects the sex organ but also hampered the functions of the accessory sex organs.

During treatment with the lindane, profound testicular changes were examined. The arrangement of Sertoli cells in the seminiferous tubules was completely damaged. This indicates that the Sertoli cells located in tubules are damaged, as effect of which sperm generation and maturation is also hampered, which was noted during studying semenology. No sperm accumulation was observed indicating dysfunction of the Sertoli cells. The interstitial space was observed to be damaged. Hence Leydig cells located in the interstitial space were degenerated, which can also be confirmed by reduction in the testosterone levels in the serum. Results obtained in this study were also reported by other articles. (52-54) A complete contradictory result was reported by Traina et al (55) indicating increase in the size of the Leydig cells due to lindane treatment. The exact mechanism of Leydig cells degeneration due to lindane treatment is yet unknown but increase in the oxidative stress is believed to be one of the major factors.

The antioxidant system plays an effective role in protecting testes and other biological tissues below a critical threshold of reactive oxygen species thus preventing testicular dysfunction. (56) Following the lindane treatment, the protein content of the tissue was observed to decline along with shrinkage of testis. The activity of the antioxidant enzymes viz. catalase and super oxide dismutase were noted to decline following the lindane treatment. The concentration of the reduced glutathione was also noted to decline in lindane treated animals. Thus the decline in the anti-oxidant parameters reveals inclination in the ROS generation in the tissue, which is also justified by inclination in the MDA concentration in the tissue. A similar result was also reported by Sujatha et al. (57) GSH, MDA and SOD levels following Quinolophos treatment and its subsequent reversal by *Withania somnifera* and *Aleo barbedensis* has also been reported in similar studies carried out on liver and kidneys. (58)

The plant extract treatment in lindane treated animals showed significant recovery. A significant recovery or restoration of the sperm and sperm function was noted at the completion of the plant extract treatment, comparable to that of the vehicle treated

control animals. The viability of the sperm and sperm density were observed to be restored, almost comparable to that of the vehicle treated control group. Sperm functional analysis parameters were also noted to be almost comparable to vehicle treated control animals.

Seminal biochemistry had revealed a significant recuperation during the plant extract treatment. The activity of LDH-C<sub>4</sub> was noted to decline compared to that of lindane treated animals, almost comparable to that of the control; revealing the improvement in the mitochondrial function as well as reduction in the oxidative stress in cauda epididymides.

ACP activity was also observed increase during the course of plant extract treatment almost comparable to that of the vehicle treated control animals group; indicating healing effect on the accessory sex organs.

The activity of the  $\beta$ -glucosidase was observed to almost equate to that of the vehicle treated control animals group at the completion of 60 days of plant extract treatment period. The result generated revealed the proper functioning of brush borders of cauda epididymides, bestowing the optimum condition for sperm storage.

Thus treatment with the plant extract reinstated the functioning of reproductive organs and accessory sex organs; almost comparable to that of control. Due to the treatment the oligo-necro-zoospermic rats were observed to produce normal sperm and fertility was observed to be restored. Thus plant extract treatment for a period of 60 days has shown ameliorative effect for damaged sperm maturation and reproductive organs functioning.

A gradual and slow re-development of testis was noticed following the plant extract treatment. On completion of the treatment, the arrangement of the Sertoli cells in the seminiferous tubules was almost comparable to that of the vehicle treated control animals. Accumulation of the sperm was also observed in the seminiferous tubules, whereas semenology studies revealed that sperm produced were duly matured indicating the proper functioning of the Sertoli cells, which were damaged during the lindane treatment. The interstitial space was also observed to re-develop following the plants extract treatment with regeneration of the Leydig cells. The levels of the testosterone in the serum were also observed to increase following the plant extract revealing a apt functioning of the Leydig cells.

During the plant extract treatment the protein content in the tissue was noted to incline along with the tissue mass, almost comparable to that of the vehicle treated animals. The activities of the antioxidant enzymes viz. catalase and superoxide dismutase were observed to decline. The

concentration of the reduced glutathione was also noted to increase during the treatment course. The concentration of the MDA was also observed to reduce. Thus plant extract treatment had diminishing effect on the oxidative stress enhanced due to lindane treatment. The antioxidant activity of the extract may be due euchrestine B, bismurrayafoline E, mahanine, mahanimbicine, and mahanimbin from *Murraya koenigii* <sup>(59)</sup> and trigonelline from *Trigonella foenum*. <sup>(24)</sup>

## CONCLUSION

Thus, in nutshell, lindane treatment has an explicit effect on reproduction. Lindane treatment induces oligo-necro-zoospermic male rats. Functioning of the reproductive organs and accessory sex organs was also hampered. Oxidative stress was also observed to increase in the tissue. Following the plant extract treatment, a significant recovery was observed. The reproductive organs functions aptly producing functionally matured sperms. The functioning of the accessory sex organs was also restored. Oxidative stress was noted to decline thus preventing the tissue from ROS damage.

## REFERENCES AND NOTES

- Vaidya A.D., 2006. Reverse pharmacological correlates of Ayurvedic drug actions. Indian J. Pharmacol. 38, 311-15.
- Patwardhan B., Vaidya A.D., Chorghade M., 2004. Ayurveda and natural products drug discovery. Curr. Sci., 86(6), 789-99.
- Vaidya A.D., Devasagayam T.P., 2007 Current status of herbal drugs in India: an overview. J. Clin. Biochem. Nutr., 41(1), 1-11.
- Reuber M.D., 1979. Carcinogenicity of lindane. Environ. Res., 19(2), 460-81.
- Xu J.K. et al., 1984. Study on the carcinogenicity of gammabenzene-hexachloride in rats. Environ. Sci. Technol., 31, 843-44.
- Kamirin M., 1997. Pesticide profiles: toxicity, environmental impact, and fate. CRC publisher, p278.
- Dalsenter P.R. et al., 1996. Reproductive toxicity and tissue concentrations of lindane in adult male rats. Hum. Exp. Toxicol., 15(5), 406-10.
- Dietrich D.R., Swenberg J.A., 1990. Lindane induces nephropathy and renal accumulation of  $\alpha$ 2u-globulin in male but not in female Fischer 344 rats or male NBR rats. Toxicol. Lett., 53(1-2), 179-81.
- Ortiz J.B., et al., 2003. Histopathological changes induced by lindane (gamma HCH) in various organs of fishes. Sci. Mart. (Barc.), 67(1), 53-61.
- Chinoy N.J., Sharma A., 1998. Amelioration of fluoride toxicity by vitamins E and D in reproductive functions of male mice. Fluoride, 31(4), 203-16.
- El-Demerdash F.M., et al., 2004. Cadmium-induced changes in lipid peroxidation, blood hematology, biochemical parameters and semen quality of male rats: protective role of vitamin E and  $\beta$ -carotene. Food Chem. Toxicol., 42(10), 1563-71.
- Santos-Filho S. et al., 2007. The male reproductive system and the effect of an extract of a medicinal plant (*Hypericum perforatum*) on the labeling process of blood constituents with technetium-99m. Braz. Arc. Biol. Techn., 50, 97-104.
- Luximon-Ramma A., et al., 2002. Antioxidant Activities of Phenolic, Proanthocyanidin, and Flavonoid Components in Extracts of *Cassia fistula*. J. Agr. Food Chem., 50(18), 5042-47.
- Iyer D., Devi P.U., 2008. Phyto-pharmacology of *Murraya koenigii* (L.). Pharmacogn. Rev., 2(3), 180-84.
- Chang F.R., et al., 2006. Anti-inflammatory and cytotoxic diterpenes from formosan *Polyalthia longifolia* var. *Pendula*. Planta Med., 73(14), 1344-47.
- Malairajan P., et al., 2008. Evaluation of anti-ulcer activity of *Polyalthia longifolia* (Sonn.) Thwaites in experimental animals. Indian J. Pharmacol., 40(3), 126-28.
- Saha M.R., et al., 2008. *In vitro* nitric oxide scavenging activity of ethanol leaf extracts of four Bangladeshi medicinal plants. Stanford J. Pharm. Sci., 1(1-2), 57-62.
- Verma M., et al., 2008. *In vitro* cytotoxic potential of *Polyalthia longifolia* on human cancer cell lines and induction of apoptosis through mitochondrial-dependent pathway in HL-60 cells. Chem. Biol. Interact., 71(1), 45-56.
- Chang H.L., et al., 2008. Inhibitory effects of 16-hydroxycyclohexa-3,13(14)E-dien-15-oic acid on superoxide anion and elastase release in human neutrophils through multiple mechanisms. Eur. J. Pharmacol., 586(1-3), 332-39.
- Islam A., et al., 2001. Antimicrobial activity and cytotoxicity of clerodane diterpenes from *Polyalthia longifolia* seeds. J. Med. Sci., 1(5), 320-23.
- Nair R., et al., 2007. Assessment of *Polyalthia longifolia* var. *Pendula* for hypoglycemic and antihyperglycemic activity. J. Clin. Diagn. Res., 1(3), 116-21.
- Abdel-Barry J.A., et al., 1997. Hypoglycaemic and antihyperglycemic effects of *Trigonella foenum-graecum* leaf in normal and alloxan induced diabetic rats. J. Ethnopharmacol., 58(3), 149-55.
- Gupta A., et al., 2001. Effect of *Trigonella foenum-graecum* (fenugreek) seeds on glycaemic control and insulin resistance in type 2 diabetes mellitus; a double blind placebo controlled study. J. Assoc. Physicians India, 49, 1057-61.
- Genet S., et al., 2002. Alterations in antioxidant enzymes and oxidative damage in experimental diabetic rat tissues: effect of vanadate and fenugreek (*Trigonella foenum graecum*). Mol. Cell Biochem., 236(1-2), 7-12.
- Vats V., et al., 2002. Evaluation of anti-hyperglycemic and hypoglycemic effect of *Trigonella foenum-graecum* Linn, *Ocimum sanctum* Linn and *Pterocarpus marsupium* Linn in normal and alloxanized diabetic rats. J. Ethnopharmacol., 79(1), 95-100.
- Raju J. et al., 2004. Diosgenin, a steroid saponin of *Trigonella foenum graecum* (Fenugreek), inhibits azoxymethane-induced aberrant crypt foci formation in F344 rats and induces apoptosis in HT-29 human colon cancer cells. Cancer Epidemiol. Biomarkers Prevent., 13(8), 1392-98.
- Kaviarasan S. et al., 2006. Fenugreek (*Trigonella foenum-graecum*) seed extract prevents ethanol-induced toxicity and apoptosis in chang liver cells. Alcohol, 41(3), 267-73.
- Kaviarasan S., Anuradha C.V., 2007. Fenugreek (*Trigonella foenum graecum*) seed polyphenols protect liver from alcohol toxicity; a role on hepatic detoxification system and apoptosis. Pharmazie., 62(4), 299-304.
- WHO laboratory manual for examination of human semen and cervical mucus interaction, 1999. Cambridge University Publisher; 27.
- Gopalkrishnan K., et al., 1994. Alteration of semen characteristics and regulatory factors in human semen with bacterial infection. Arch. Andro., 32(3), 213-18.
- Gopalkrishnan K., et al., 1991. *In vitro* decondensation of nuclear chromatin of human spermatozoa; assessing fertilizing potential. Arch. Androl., 27(1), 43-50.
- Jeyendran R.S., et al., 1984. Development of an assay to assess the functional integrity of the human sperm membrane and its relationship to other semen characteristics. J. Reprod. Fertil., 70(1), 219-28.
- Mihara M., Uchiyama M., 1978. Determination of malonaldehyde precursor in tissue using thiobarbituric acid test. Anal. Biochem., 86(1), 271-78.
- Beutler E., Yeh M.K., 1963. Erythrocyte Glutathione Reductase. Blood, 21, 573-85.
- Tarpey M.M., Fridovich I., 2001. Methods of detection of vascular reactive species nitric oxide, superoxide, hydrogen peroxide, and peroxynitrite. Circ. Res., 89, 224-36.
- Aebi H., 1984. Catalase *in vivo*. Method Enzymol., 105, 121-26.
- Shivanandappa T., Krishnakumari M.K., 1983. Hexachlorocyclohexane induced testicular dysfunction in rats. Acta Pharmacol. Toxicol., 52(1), 12-17.
- Roy Chowdhury A., et al., 1987. Testicular changes of rat under lindane treatment. B Environ. Contam. Toxicol., 38(1), 154-56.
- Prasad A.K., et al., 1995. Effect of dermal application of hexachlorocyclohexane (HCH) on male reproductive system of rat. Hum. Exp. Toxicol., 14(6), 484-88.
- Dalsenter P.R., et al., 1997a. Serum testosterone and sexual behaviour in rats after prenatal exposure to lindane. B Environ. Contam. Toxicol., 59(3), 360-66.
- Dalsenter P.R., et al., 1997b. Reproductive toxicity and toxicokinetics of lindane in the male offspring of rats exposed during lactation. Hum. Exp. Toxicol., 16(3), 146-53.
- Beard A.P., et al., 1999. Reproductive and endocrine function in rams exposed to the organochlorine pesticides lindane and pentachlorophenol from conception. J. Reprod. Fertil., 115(2), 303-14.
- Saradha B., Mathur P.P., 2006. Induction of oxidative stress by lindane in epididymis of adult male rats. Environ. Toxicol. Phar., 22(1), 90-96.
- Beard A.P., Rawlins N.C., 1998. Reproductive effects in mink (*Mustela vison*) exposed to the pesticides lindane, carbofuran and pentachlorophenol in a multigeneration study. J. Reprod. Fertil., 113, 95-104.
- Fausto A.M., et al., 2001. Sperm quality and reproductive traits in male offspring of female rabbits exposed to Lindane (gamma-HCH) during pregnancy and lactation. Reprod. Nutr. Dev., 41(3), 217-25.
- Samanta L., Chainy G.B., 1997. Comparison of hexachlorocyclohexane-induced oxidative stress

- in the testis of immature and adult rats. *Comp. Biochem. Phys. C*, 118(3), 319-27.
47. Sawane M.V., et al., 2002. Seminal LDH-C<sub>4</sub> isoenzyme and sperm mitochondrial activity; a study in male partners of infertile couples. *Indian J. Med. Sci.*, 56(11), 560-61.
48. Ando S., et al., 1990. Fructose, prostatic acid phosphatase and zinc levels in the seminal plasma of varicoceles. *Int. J. Fertil.*, 35(4), 249-52.
49. Kalla N.R., et al., 1997. Alpha-glucosidase activity in the rat epididymis under different physiological conditions. *Int. J. Androl.*, 20(2), 92-95.
50. Yeung C.H., Cooper T.G., 1994. Study of the role of epididymal  $\alpha$ -glucosidase in the fertility of male rats by the administration of the enzyme inhibitor castanospermine. *J. Reprod. Fertil.*, 102(2), 401-10.
51. Guerin J.F., et al., 1990. Seminal alpha-glucosidase activity as a marker of epididymal pathology in nonazoospermic men consulting for infertility. *J. Androl.*, 11(3), 240-45
52. Suwalsky M., et al., 2000. Plasma absorption and ultrastructural changes of rat testicular cells induced by lindane. *Hum. Exp. Toxicol.*, 19(9), 529-33.
53. Ronco A.M., et al., 2001. The mechanism for lindane-induced inhibition of steroidogenesis in cultured rat Leydig cells. *Toxicol.*, 159(1-2), 99-106.
54. Ananya R., et al., 2005. Oxidative stress and histopathological changes in the heart following oral lindane (gamma hexachlorohexane) administration in rats. *Med. Sci. Monit.*, 11(9), BR325-329.
55. Traina M.E., et al., 2003. Long-lasting effects of lindane on mouse spermatogenesis induced by *in utero* exposure. *Reprod. Toxicol.*, 17(1), 25-35.
56. Oschendorf F.R., 1999. Infections in the male genital tract and reactive oxygen species. *Hum. Reprod. Update*, 5(5), 399-420.
57. Sujatha R., et al., 2001. Effect of lindane on testicular antioxidant system and steroidogenic enzymes in adult rats. *Asian J. Androl.*, 3(2), 135-38.
58. Surana B., et al., 2009. Toxicological effects of QUinolphos and its subsequent reversal by using root extract of *Withania somnifera* and leaf pulp of *Aloe barbadensis*. *J. Indian Soc. Toxicol.*, 4(2), 1-5.
59. Tachibana Y., et al., 2001. Antioxidative activity of carbazoles from *Murraya koenigii* leaves. *J. Agri. Food Chem.*, 49(11), 5589-94.

**Acknowledgement:** The authors are grateful for the financial assistance from Nirma Education & Research Foundation (NERF), Ahmedabad for the present investigation. The present investigation is a part the Dissertation Thesis of Mr. Ankit Kumar Jain and Mr. Bandish Kapadia submitted to Nirma University, Ahmedabad.